

A New Theranostic System Based on Gold Nanocages and Phase-Change Materials with Unique Features for Photoacoustic Imaging and Controlled Release

Geon Dae Moon,^{†,‡} Sung-Wook Choi,^{†,§} Xin Cai,[†] Weiyang Li,[†] Eun Chul Cho,[†] Unyong Jeong,[‡] Lihong V. Wang,[†] and Younan Xia^{*,†}

[†]Department of Biomedical Engineering, Washington University, St. Louis, Missouri 63130, United States [‡]Department of Materials Science and Engineering, Yonsei University, 134 Shinchon-dong, Seoul, Korea

S Supporting Information

ABSTRACT: This communication reports a new theranostic system with a combination of capabilities to both enhance the contrast of photoacoustic (PA) imaging and control the release of a chemical or biological effector by high-intensity focused ultrasound (HIFU). The fabrication of this system simply involves filling the hollow interiors of gold nanocages with a phase-change material (PCM) such as 1-tetradecanol that has a melting point of 38–39 °C. The PCM can be premixed and thus loaded with a dye, as well as other chemical or biological effectors. When exposed to direct heating or HIFU, the PCM will melt and escape from the interiors of nanocages through small pores on the surface, concurrently releasing the encapsulated molecules into the surrounding medium. We can control the release profile by varying the power of HIFU, the duration of exposure to HIFU, or both.

ollow nanostructures have been extensively explored as Hdrug delivery systems for biomedical applications due to their unique capability to hold and release drugs.¹ Most of these structures are composed of lipids, including liposomes (uni- and multilamella vesicles) and multichanneled cubo- or hexosomes.² Polymeric nanoparticles with various compositions, structures, and porosities have also been investigated for similar applications.³ Typically, the interiors of these hollow nanostructures are loaded with drugs (hydrophilic or hydrophobic) in different formulations that are able to escape via diffusion or degradation-triggered release. In addition, the release can be triggered and regulated in response to environmental changes such as pH and temperature. Recently, inorganic and composite hollow nanostructures have also received attention for drug delivery applications. A number of groups have reported delivery systems based on mesoporous silica particles containing water-insoluble drugs,⁴ and metal-organic framework (MOF) particles incorporated with drugs.⁵ Although the inclusion of inorganic components such as metal ions may cause potential toxicity issues, it offers advantages such as multifunctionality, a feature required for future theranostic applications. For example, the metal ions can serve as contrast agents for magnetic resonance imaging (MRI), adding a unique capability to resolve and track the in vivo location of a drug delivery system.⁶

Hollow nanostructures of noble metals, such as Au nanocages (AuNCs), represent another attractive platform for theranostic applications. Thanks to their strong, highly tunable scattering and absorption in the near-infrared (NIR) from 700 to 900 nm, AuNCs are wonderful contrast agents for a variety of imaging modalities including photoacoustic tomography (PAT) and optical coherence tomography (OCT).7 They have also been used to control the release of hydrophilic drugs with NIR laser or HIFU by coating their surfaces with smart polymers.⁸ While the NIR laser-triggered release has to rely on the photothermal effect of AuNCs, HIFU can directly deposit acoustic energy in the focal volume to rapidly raise the temperature.^{8b} The availability of AuNCs loaded with therapeutic drugs offers a great benefit to theranostic applications because the AuNCs can be monitored with an optical imaging technique while the drug is released at the targeted site in a controllable fashion. As expected, the released drug can also greatly enhance the efficacy of photothermal cancer treatment, which has been demonstrated with success in the absence of an anticancer drug.⁹ While the system based on AuNCs and smart polymers has been successfully used for controlled release under the irradiation of NIR laser or HIFU, the loaded drugs can escape from the hollow interiors of AuNCs through a slow diffusion process in the absence of any thermal stimulus. In addition, this system only works for hydrophilic drugs because the as-prepared nanocages are typically coated with a hydrophilic polymer such as poly(vinyl pyrrolidone), making it difficult for a hydrophobic species to diffuse into the hydrophilic interior of a AuNC. Since more than 40% of the active compounds identified by the combinatorial screening programs are poorly soluble in water,¹¹ finding a system well-suited for encapsulation of both hydrophilic and hydrophobic drugs is expected to greatly extend their scope of use in theranostic applications. In addition to the features needed for imaging purposes, the new system should show negligible release until it is triggered by an external stimulus.

Herein, we present a facile and versatile strategy for loading either hydrophobic or hydrophilic drugs into the hollow interiors of AuNCs. We use a phase-change material (PCM) as the medium to help load the drug, which can also serve as a "gatekeeper" to control the release of drug in response to temperature increase.¹¹ In principle, the encapsulated drug should not be released until the PCM has been melted due to heating by a



Received:January 28, 2011Published:March 14, 2011

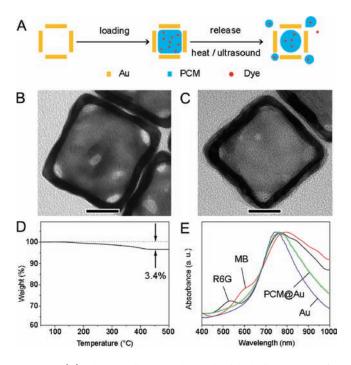


Figure 1. (A) Schematic illustrating how to load the hollow interior of a AuNC with a dye-doped PCM and then have it released from the AuNC by direct or ultrasonic heating. (B, C) Typical TEM images of a AuNC before and after its hollow interior had been loaded with a mixture of 1-tetradecanol (the PCM) and R6G (the drug). The scale bars are 20 nm. (D) TGA data of the pristine AuNCs (dotted) and the AuNCs containing PCM/R6G (solid). (E) UV–vis absorption spectra of the AuNCs before and after loading with the PCM/dye mixtures.

thermal, photothermal, or ultrasonic means. Figure 1A shows a schematic diagram of the encapsulation and release mechanisms. Since a PCM reversibly changes its physical states between solid and liquid over a narrow temperature range, it can perfectly confine drug molecules inside the AuNCs at a temperature below its melting point (the solid state). When the local temperature is raised beyond the melting point of the PCM, it will begin to melt (the liquid state), and the drug will be released from the melted PCM through diffusion. Taken together, this new system should allow regulating the release of a drug and the profile by manipulating the temperature. As long as the drug is miscible with the PCM phase, it can be conveniently loaded into the hollow interiors of AuNCs as the PCM diffuses into the nanocages. This requirement can be readily met by choosing PCMs with surfactantlike behavior, such as those containing both long hydrophobic tails and hydrophilic heads. In this work, we chose to focus on 1-tetradecanol, a fatty alcohol characterized by attractive features such as immiscibility with water, good biocompatibility, and a melting point $(38-39 \degree C)$ slightly higher than the normal human body temperature $(37.0 \degree C)$. It can also be mixed with a range of hydrophilic and hydrophobic substances. Furthermore, 1-tetradecanol is an ingredient widely used in cosmetics due to its low toxicity (oral, rat $LD_{50} > 5 g/kg$).¹²

The AuNCs used in this study were prepared by means of a galvanic replacement reaction between Ag nanocubes (44 \pm 6.7 nm in edge length) and HAuCl₄ in an aqueous solution under refluxing condition.^{13b} The resultant AuNCs had an outer edge length of 60 \pm 11 nm, together with a wall thickness of 7.5 \pm 2.8 nm. Essentially all of them contained at least one pores (~10 nm in size) on the surface (Figure S1, Supporting Information

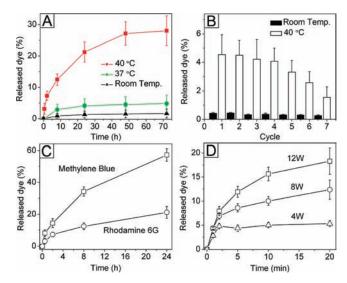


Figure 2. Release profiles of (A) R6G under direct heating to various temperatures for different periods of time, (B) R6G through different cycles of heating (40 $^{\circ}$ C, for 2 h) and cooling (to room temperature), (C) R6G and MB by direct heating to 40 $^{\circ}$ C for different periods of time, and (D) R6G by HIFU at different applied powers.

[SI]). The as-prepared nanocages were then redispersed in methanol after repeated washing with deionized water and centrifugation 4 times. For encapsulation, we used Rhodamine 6G (R6G) and methylene blue (MB) as two examples of drugs with different solubilities in water. As a major advantage over most real drugs, the release of these two dyes can be easily monitored and quantified through the use of UV—vis absorption spectroscopy.

In a typical experiment, the PCM was added to a glass vial and placed in an oil bath set to 90 °C to melt the PCM into a liquid, followed by the addition of a dye. After the dye and PCM had been thoroughly mixed, we introduced the AuNCs as a suspension in methanol. Even after the methanol had been evaporated due to stirring and heating, the AuNCs were still well dispersed in the liquid PCM. During this process, the mixture of PCM and dye molecules slowly entered the hollow interior of each nanocage by diffusion through the pores on the surface. After the system had continued being heated at 90 °C for 2 h, a small amount of hot water was added to generate two separated phases, one being the PCM/ dye mixture and the other containing the loaded AuNCs and water, as a result of immiscibility between the PCM and water. The AuNCs were preferentially extracted into the water phase due to their hydrophilic surfaces. Interestingly, both PCM and dye molecules inside the nanocages were effectively retained because it was difficult for the PCM molecules to quickly diffuse into water due to their immiscibility. Finally, the AuNCs encapsulated with the PCM/dye mixture were collected by centrifugation and then redispersed in deionized water under brief sonication (for 10 s). In the course of redispersion, a very small portion of the encapsulated PCM/dye could escape from the AuNCs owing to the heat generated by sonication (Figure S2, SI).

Panels B and C of Figure 1 respectively show typical transmission electron microscopy (TEM) images of the nanocages before and after loading of the PCM/dye mixture. Compared with the pristine sample, nanocages loaded with the PCM/dye mixture showed a conspicuous difference in contrast. It should be pointed out that the PCM/dye mixture inside the nanocage was often smaller in volume than the actual void of the nanocage. To quantify the amount of PCM/dye loaded inside the AuNCs, we obtained thermal gravimetric analysis (TGA) data in the range of 50-500 °C (Figure 1D). The TGA results show a weight loss of 3.4%, which is lower than the loading of 4.0% calculated from the density of PCM and physical dimensions of the nanocages (SI). These data suggest that, on average, only 84% of the void space inside each AuNC was actually occupied by the PCM/dye mixture.

Figure 1E shows UV—vis absorption spectra recorded from aqueous suspensions of pristine AuNCs, AuNCs containing the PCM, and AuNCs containing both PCM and the dye (R6G or MB). It is worth pointing out that the AuNCs loaded with a PCM/dye mixture also exhibited the characteristic peak of the dye, confirming successful encapsulation of the dye inside the AuNCs. The major localized surface plasmon resonance (LSPR) peak of the nanocages was found to be broadened and slightly red-shifted, most likely caused by some minor aggregation during sample preparation.

Figure 2 shows the release of R6G from the AuNCs by direct heating and HIFU heating, respectively. As shown in Figure 2A, the amount of R6G released was negligible at room temperature because the dye was entrapped by solid PCM inside the nanocages, which makes it difficult for the dye molecules to diffuse into the surrounding medium. At 37 °C, there was a slight increase in the release of R6G (still below 5% over a period of 3 days) although the melting point of 1-tetradecanol $(38-39 \degree C)$ was still slightly higher than this temperature. In comparison, 28% of the encapsulated R6G was released from the AuNCs in 3 days when the sample was held at 40 °C. As an important feature, the PCM-based release could be easily regulated by controlling the temperature. We confirmed this feature by investigating the release behaviors during repeated heating-cooling cycles at temperatures below and above the melting point of 1-tetradecanol (Figure 2B). Note that the release of dye was only observed when the sample was heated to 40 $^{\circ}$ C and held for 2 h.

Due to its good compatibility with both hydrophobic and hydrophilic substances, we found that the PCM could also be used as a matrix for the encapsulation and release of a drug more soluble in water than R6G is. We demonstrated this capability by switching to MB, a dye more hydrophilic than R6G. In this case, we found that the loading capacity of MB was much lower than that of R6G (57% vs 84%, respectively, see SI). As a result, the cumulative release of MB was 3 times higher than that of R6G (Figure 2C). Since MB was quickly photobleached (Figure S3, SI), the cumulative release percentage corrected for bleaching showed that 80% of the loaded MB was released in 24 h (Figure S4, SI). These results clearly demonstrate that the PCM works for chemical species with different solubilities in water, making this new system a useful platform for a wide variety of drugs.

Figure 2D shows the release profiles of R6G as a function of the power of HIFU. HIFU has been used as a promising clinical technology for tumor treatment due to its deep penetration and noninvasive nature.¹⁴ Most recently, we have also adapted this technique for triggering the release of dye molecules from AuNCs whose surfaces were covered with smart polymer brushes.^{8b} As a major advantage over the smart polymers, the PCM-based system showed better encapsulation efficacy and much lower background release (data not shown). In the present work, we also found that the focused ultrasound wave could rapidly increase the overall temperature of a suspension of AuNCs loaded with PCM and dye (Figure S5, SI). As clearly shown in Figure 2D, the quick increase in temperature caused the R6G to release from the hollow interiors of AuNCs. As expected, the release profile also displayed a strong dependence on the power of HIFU, making it possible to control the release dosage on demand.

In conclusion, we have demonstrated a nanoscale, temperature-regulated drug release system by combining the unique features of AuNCs and PCMs. As a deeply penetrating energy source, HIFU can be used to trigger the release and to control the release profile by adjusting the duration of exposure and/or the power applied. The new hybrid system described in this work can also be further developed into a theranostic system with an array of functions, including the capabilities for in vivo molecular imaging, as well as chemo- and photothermal therapy.

ASSOCIATED CONTENT

Supporting Information. Synthesis procedures and other supplementary data. This material is available free of charge via the Internet at http://pubs.acs.org.

AUTHOR INFORMATION

Corresponding Author xia@biomed.wustl.edu

Present Addresses

[§]Division of Biotechnology, The Catholic University of Korea, Gyeonggi-do, Korea

ACKNOWLEDGMENT

This work was supported in part by a 2006 NIH Director's Pioneer Award (DP1 OD000798), startup funds from Washington University in St. Louis, and the World Class University (WCU) program at Yonsei University. Part of the work was done at the Nano Research Facility (NRF), a member of the National Nanotechnology Infrastructure Network (NINN), which is supported by the NSF under Award No. ECS-0335765.

REFERENCES

(1) (a) Lou, X. W.; Archer, L. A.; Yang, Z. Adv. Mater. 2008, 20, 3987. (b) Li, Z.-Z.; Wen, L.-X.; Shao, L.; Chen, J.-F. J. Controlled Release 2004, 98, 245.

(2) (a) Lasic, D. D. Trends Biotechnol. 1998, 16, 307. (b) Drummond,
D. C.; Meyer, O.; Hong, K.; Kirpotin, D. B.; Paapahadjopoulos, D.
Pharmacol. Rev. 1999, 51, 691. (c) Zamboni, W. C. Oncologist 2008, 13, 248.

(3) (a) Nasongkla, N.; Be, E.; Ren, J.; Ai, H.; Khemtong, C.; Guthi, J. S.; Chin, S.-F.; Sherry, A. D.; Bothman, D. A.; Gao, J. *Nano Lett.* **2006**, *6*, 2427. (b) Medina, S. H.; El-Sayed, M. E. H. *Chem. Rev.* **2009**, *109*, 3141. (c) De Cock, L. J.; De Koker, S.; De Geest, B. G.; Grooten, J.; Vervaet, C.; Remon, J. P.; Sjihorukov, G. B.; Antipina, M. N. *Angew. Chem., Int. Ed.* **2010**, *49*, 2. (d) Tong, R.; Christian, D. A.; Tang, L.; Cabral, H.; Baker, J. R., Jr.; Kataoka, K.; Discher, D.; Cheng, J. *MRS Bull.* **2009**, *34*, 422.

(4) (a) Rosenholm, J. M.; Peuhu, E.; Eriksson, J. E.; Sahlgren, C.; Linden, M. *Nano Lett.* **2009**, *9*, 3308. (b) Wang, Y.; Yan, Y.; Cui, J.; Hosta-Rigau, L.; Heath, J. L.; Nnice, E. C.; Caruso, F. *Adv. Mater.* **2010**, *22*, 4293.

(5) (a) Horcajada, P.; Serre, C.; Vallet-Regi, M.; Sebban, M.; Taulelle, F.; Ferey, G. Angew. Chem., Int. Ed. 2006, 45, 5974. (b) Uemura, K.; Kitagawa, S.; Kondo, M.; Fukui, K.; Kitaura, R.; Chang, H.-C.; Mizutani, T. Chem.-Eur. J. 2002, 8, 3586.

(6) (a) Lin, W.; Hyeon, T.; Lanza, G. M.; Zhang, M.; Meade, T. J. *MRS Bull.* **2009**, *34*, 441.(b) Que, E. L.; Chang, C. J. *Chem. Soc. Rev.* **2010**, *39*, 51. (c) Della Rocca, J.; Lin, W. *Eur. J. Inorg. Chem.* **2010**, 3725.

Journal of the American Chemical Society

(7) (a) Kim, C.; Cho, E. C.; Chen, J.; Song, K. H.; Au, L.; Favazza, C.;
Zhang, Q.; Cobley, C. M.; Gao, F.; Xia, Y.; Wang, L. V. ACS Nano 2010,
4, 4559. (b) Yang, X.; Skrabalak, S.; Li, Z.; Xia, Y.; Wang, L. V. Nano Lett.
2007, 7, 3798. (c) Song, K. H.; Kim, C.; Cobley, C. M.; Xia, Y.; Wang,
L. V. Nano Lett. 2009, 9, 183. (d) Chang, H.; Sun, T.; Li, Z. Y.; Chen, J.;
Wiley, b. J.; Xia, Y.; Li, X. Opt. Lett. 2005, 30, 3048.

(8) (a) Mustafa, S. Y.; Cheng, Y.; Chen, J.; Cobley, C. M.; Zhang, Q.; Rycenga, M.; Xie, J.; Kim, C.; Song, K. H.; Schwartz, A. G.; Wang, L. V.; Xia, Y. *Nat. Mater.* **2009**, *8*, 935.(b) Li, W.; Cai, X.; Kim, C.; Sun, G.; Zhang, Y.; Deng, R.; Yang, M.; Chen, J.; Achilefu, S.; Wang, L. V.; Xia, Y. *Nanoscale* **2011**, 10.1039/C0NR00932F.

(9) (a) Skrabalak, S. E.; Chen, J.; Au, L.; Lu, X.; Li, X.; Xia, Y. Adv. Mater. 2007, 19, 3177. (b) Chen, J.; Wang, D.; Xi, J.; Au, L.; Siekkinen, A.; Warsen, A.; Li, Z.-Y.; Zhang, H.; Xia, Y.; Li, X. Nano Lett. 2007, 7, 1318. (c) Chen, J.; Glaus, C.; Laforest, R.; Zhang, Q.; Yang, M.; Gidding, M.; Welch, M. J.; Xia, Y. Small 2010, 6, 811.

(10) (a) Lipinski, C. A. J. Pharmacol. Toxicol. Methods 2000, 44, 235.
(b) Merisko-Liversidge, E. M.; Liversidge, G. G. Toxicol. Pathol. 2008, 36, 43.

(11) (a) Mondal, S. Appl. Therm. Eng. 2007, 28, 1536. (b) Choi, S.-W.; Zhang, Y.; Xia, Y. Angew. Chem., Int. Ed. 2010, 49, 7904.

(12) (a) Gosselin, R. E.; Hodge, H. C.; Smith, R. P.; Gleason, M. N. *Clinical Toxicology of Commercial Products*, 4th ed.; Williams and Wilkins: Baltimore, 1976; pp II–118. (b) Material Safety Data Sheet: MSDS ALCH 414-1.

(13) (a) Zhang, Q.; Li, W.; Wen, L.-P.; Chen, J.; Xia, Y. *Chem.-Eur. J.* **2010**, *16*, 10234. (b) Skrabalak, S. E.; Chen, J.; Sun, Y.; Lu, X.; Au, L.; Cobley, C. M.; Xia, Y. *Acc. Chem. Res.* **2008**, *41*, 1587.

(14) Kennedy, J. E.; ter Haar, G. R.; Cranston, D. *Br. J. Radiol.* 2003, 76, 590.